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Abstract

Since long-term exposure of cancer-related therapeutics have been linked to alterations of cardiac function in patients, the Stem Cell Working Group as part of the Health and Environmental Science Institute (HESI) endeavors to gain further insight into chronic cardiotoxicity. The objective of the study was to optimize non-clinical safety assessment strategies of chronic cardiotoxicity by testing prolonged exposure of reference compounds on cell-based assay systems using human induced pluripotent stem cell-derived cardiomyocytes (hiPSC-CMs). We analyzed the effects of compounds with known cardiotoxic potential and diverse mechanisms of action (MoAs) on contractile and electrophysiological properties of hiPSC-CMs over a period of up to 120 hours. This excerpt of the HESI Stem Cell Working Group study underlines the potential of *in vitro* systems to address contractile function of hiPSC-CMs for chronic safety pharmacological studies of compounds with diverse MoAs.

Study Outline

A panel of 12 compounds was selected to evaluate chronic adverse side effects on hiPSC-CMs, including alterations in energetics and contraction force as well as electrophysiological and structural disturbance.

For the quantification of adverse side effects, a variety of methods was used, covering electrophysiology, mechanobiology, optical methods as well as biochemical assays.

Target Toxicity	Compound	Conc. Range	Parameter	Method
Energetics/ Mitochondrial Toxicity	Doxorubicin	0.1 - 3 µM	Electrophysiology	Multielectrode array
	Erlotinib	0.3 - 10 µM		Impedance
	Sunitib	0.01 - 1 µM	Mechanobiology	Contraction force
Electrophysiological Disturbance	Pentamidine	0.1 - 3 µM	Optical Methods	Optical contraction analysis
	Arsenic Trioxide	0.1 - 3 µM		Voltage sensitive dye
Contractility	BMS-986094	0.1 - 3 µM		Calcium imaging
	Milrinone	0.1 - 10 µM	Biochemical	Oxygen consumption rate
	Nilotinib	0.1 - 10 µM		Biomarkers
Structural/myofilament Disturbance	Endothelin-1	0.3 - 100 nM		
	Vinblastine	1 - 300 nM		
	Vincristine	1 - 300 nM		
	Vinorelbine	0.1 - 3 µM		

green: presented in this poster

↑ **Table 2:** Methods used for the quantification of chronic cardiotoxic compound effects

← **Table 1:** Cardiotoxic compounds sorted by target toxicity

Workflow

hiPSC-CMs were seeded from cryopreserved state into the assay format and cultured in maintenance medium for 5 days. 24h prior to compound application, medium was changed to serum-free, protein-containing assay medium. Baseline measurements were performed before compound application. Compounds were added 4x concentrated during a partial media change. The first compound measurement was carried out after 1h. Subsequently, chronic compound responses were recorded in 24h intervals for a total of 120h.

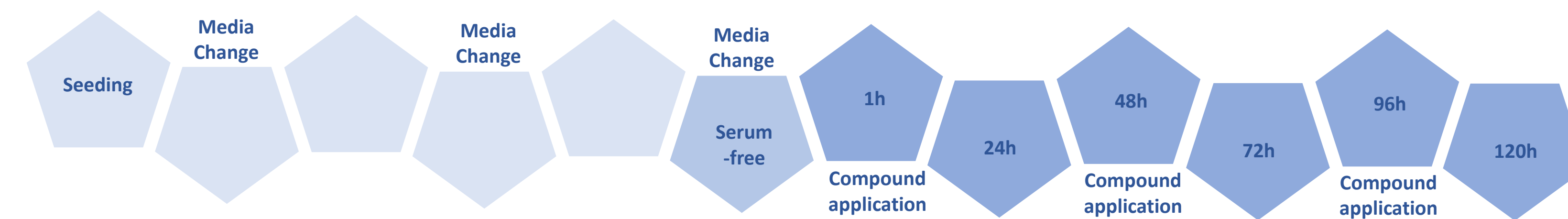


Figure 2: Study workflow

Results

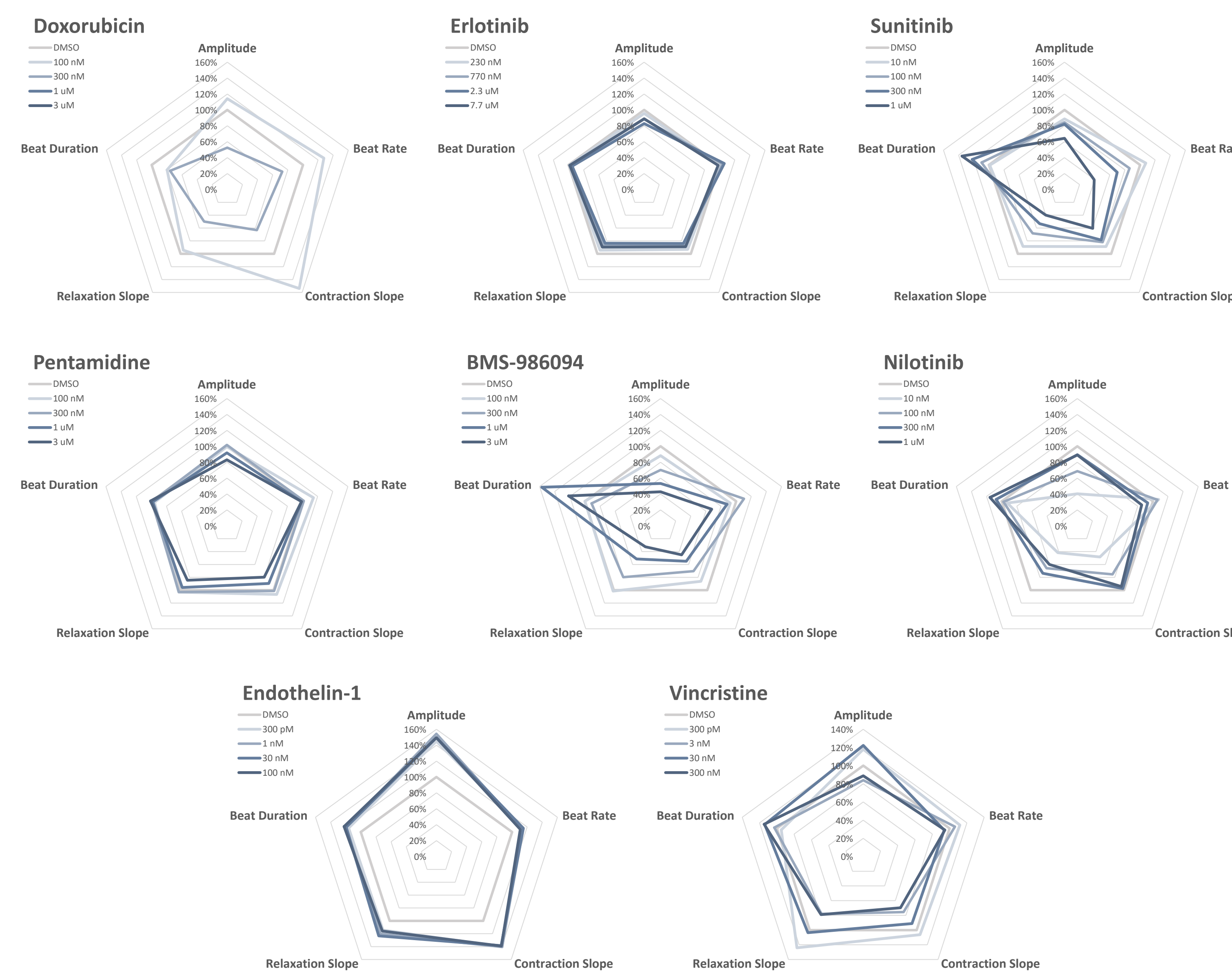


Figure 3: Contraction parameter analysis at 72h post compound application. Parameters: Contraction force (Amplitude), beat rate, contraction slope, relaxation slope, beat duration

Target Toxicity	Compound	C1	C2	C3	C4
Energetics/ Mitochondrial Toxicity	Doxorubicin	0.75	0.45	0.40	0.36
	Erlotinib	0.95	0.99	1.03	1.00
	Sunitinib	0.94	1.10	1.23	1.11
Ephys Disturbance	Pentamidine	0.94	0.94	0.81	0.62
	BMS-986094	0.93	0.78	0.83	0.77
Contractility	Nilotinib	0.96	1.09	1.14	1.00
Structural/myofilament Disturbance	Endothelin-1	0.82	0.83	0.86	0.87
	Vincristine	0.90	0.57	0.43	0.46

Table 3: Effect of the tool compounds on the base impedance as a measure of monolayer integrity

Technology

iCell Cardiomyocytes² are highly pure human iPSC-derived cardiomyocytes and allow for robust and reproducible quantification of compound effects as demonstrated in the CiPA study (Blinova *et al.*, 2017).

The **CardioExcyte 96** is an impedance-based system where hiPSC-derived cardiomyocytes are cultured on gold electrodes, enabling label-free and continuous quantification of electrical impedance as a measure of monolayer integrity.

In the **FLEXcyte 96** system, hiPSC-derived cardiomyocytes are cultured on ultra-thin silicone membranes, mimicking the mechanical environment of native human heart tissue. Rhythmic contractions deflect the membranes, quantified by capacitive distance sensing. Parameters analyzed are contractile force (AMP), beat rate (BR), beat rate regularity (BRR), rising time (RT), falling time (FT). (Gossmann *et al.*, 2016 and 2020).

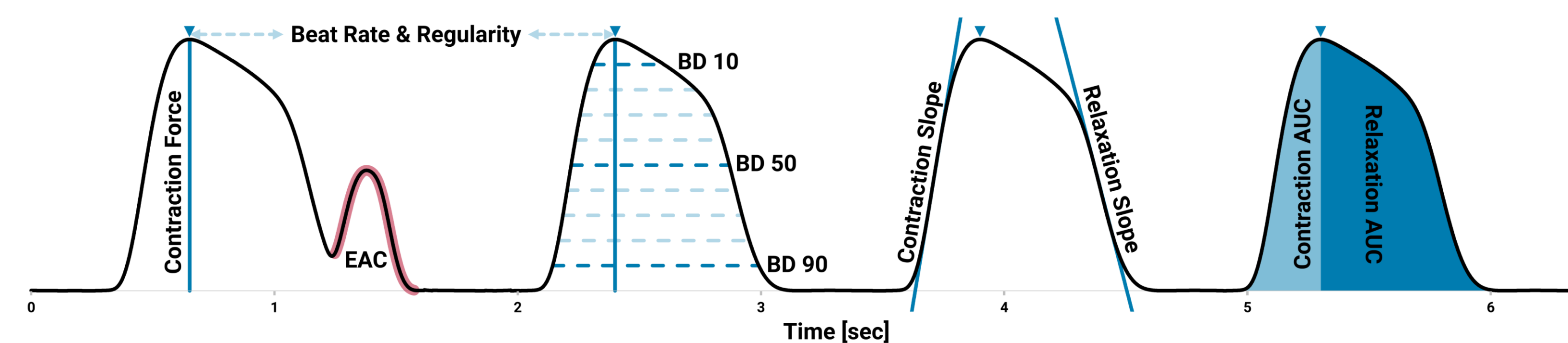


Figure 1: FLEXcyte 96 contraction analysis parameters

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